Effects of obesity on liver cytochromes P450 in various animal models

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The prevalence of obesity and other obesity-related diseases is increasing worldwide. Obesity is a disease characterized by increased body weight, or a condition resulting from excessive accumulation of body fat. Due to increased body fat deposits, obesity has also been associated with increased mortality resulting from higher incidence rates of hypertension, diabetes, or various types of cancer, such as breast, colorectal, cervical and prostate cancer. Physiological changes associated with obesity are likely to result in altered drug biotransformation. The main enzymes enabling the oxidative biotransformation of most drugs are cytochromes P450 (CYPs). The review summarizes how pathophysiological factors, especially obesity, affect properties (e.g. enzyme activity, protein expression, gene expression) of CYP enzymes in various experimental models of human obesity. Results reported by various authors suggest that obesity is associated with a decrease of CYP activities (except for the CYP2C and CYP2E1 enzymes). The only exception is mouse obesity induced by monosodium glutamate (administered to newborn mice) as it usually leads to increased CYP expression. Selecting an animal model that is as close as possible to the properties of human obesity is of paramount importance.

Key words: obesity, animal model, cytochrome P450, drug metabolism, enzyme activity, protein level, mRNA level

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INTRODUCTION

Obesity is a serious metabolic disorder that has become a global health problem. It is often considered a low-grade inflammatory condition¹ resulting in several chronic human diseases²⁻⁴ including hypertension, diabetes⁵ and various types of cancer, such as breast, endometrial, colorectal, cervical, ovarian, stomach and prostate cancer^{3,6}. As mentioned previously, obesity results from excessive accumulation of body fat⁵. To classify the overweight status and obesity in adults, the body mass index (BMI) is commonly used that is calculated by dividing weight (in kilograms) by the square of height (in meters). Overweight occurs when BMI \geq 25, obesity is defined as BMI \geq 30 (ref.⁷). According to the World Health Organization, around two billion adults aged 18 years and above were overweight in 2014. Of these, more than 600 million were obese. If the prevalence of obesity continues to rise linearly, 51% of the population will become obese in 2030 (ref.8).

Physiological changes associated with obesity are also likely to involve altered drug metabolism and clearance of drugs^{2,9}. These may lead to an increased risk of adverse drug effects and drug interactions in obese individuals. Therefore, it is very important to know whether enzymes involved in biotransformation of drugs are influenced by pathological conditions in individuals with obesity^{5,9,10}; animal models are commonly used to investigate the corresponding consequences.

DRUG-METABOLIZING ENZYMES

Drug metabolism is mainly carried out by drug-metabolizing enzymes which correspond to two phases of biotransformation (phase I and phase II). Cytochrome P450 (CYP) enzymes belong to phase I of biotransformation and represent a superfamily of heme-containing enzymes^{9,11}. Phase I of biotransformation of foreign compounds includes oxidation, reduction or hydrolysis converting the parent substance to a more polar metabolite¹². Almost sixty CYP enzymes have been found in the human genome. CYP enzymes are essential for detoxification of foreign chemicals (xenobiotics) and for metabolism of many medications^{13,14}. Human CYP enzymes participate in the biotransformation of most xenobiotics including more than two thirds of all medications in clinical use. The most important CYP enzymes participating in drug metabolism include CYP1A1, CYP1A2, CYP2A6, CYP2B6, CYP2C8, CYP2C9, CYP2C19, CYP2D6, CYP2E1, CYP3A4 and CYP3A5 (ref. 13,15). Most CYP enzymes in the human organism are primarily expressed in the liver¹³, but they are also present in the small intestine, heart, lungs, placenta, kidneys and other organs¹⁴. The most common form expressed in the human liver which metabolizes almost one half of marketed drugs is CYP3A4 (ref.¹³).

In the following subsections, the most important CYP enzymes participating in drug metabolism are presented. Also, the results of animal and/or human studies that

describe the effects of pathophysiological factors, particularly obesity, on the properties of CYP enzymes are discussed.

Subfamily of CYP1A enzymes

The human CYP1A subfamily includes two main members, CYP1A1 and CYP1A2 (ref.¹¹). The CYP1A1 form is predominantly expressed extrahepatically; in the liver, it is expressed as a result of induction, for example by polycyclic aromatic compounds. On the contrary, the CYP1A2 form is mostly expressed in the human liver¹³ and either not or only weakly expressed in extrahepatic tissues¹⁶. CYP1A2 plays a more important role in drug metabolism than CYP1A1 (ref.¹⁷). Human hepatic CYP1A2 accounts for approximately 13% of total hepatic CYP content and is responsible for metabolism of ~10% of marketed drugs^{13,16}. Various substrates are metabolized via the CYP1A2 enzyme, including caffeine¹⁸ and the ophylline⁹.

Metabolism of specific probes of CYP1A2 activity has been studied in obese in comparison with non-obese populations¹⁹. A recent study focused on determining the oxidative enzyme activity of CYP1A2 in non-obese and obese children using caffeine as a probe substrate. The authors found no changes in CYP1A2 activity in urine samples between non-obese and obese children¹⁸.

Similarly, no changes or no significant changes were found in two animal studies. An *in vivo* study showed no significant differences in CYP1A activity when comparing lean and *ob/ob* mice; ethoxyresorufin was used as a probe for CYP1A activity²⁰. The other study revealed unaltered expression of *Cvp1a2* in high-fat diet mice⁹.

Yoshinari et al.²¹ provided data showing that mRNA levels of the *Cyp1a2* gene did not differ between *db/db* mice and control animals. However, protein expression of CYP1A2 tended to decrease in *db/db* mice. Also in our recent study¹⁰ of obese mice, no tendency to reduction of mRNA and protein expression was detected. On the contrary, significantly increased CYP1A2 activity in monosodium glutamate (MSG)-obese mice was found.

Obese individuals are more susceptible to cancers of the breast, cervix, ovary and uterus. 7,12-Dimethylbenz(a) anthracene is used as a model compound for inducing mammary carcinogenesis in rodents. Hakkak et al.²² used 40-day-old ovariectomized obese and non-obese Zucker diabetic fatty (ZDF) rats. The authors observed significantly higher protein expression of CYP1A2 in the liver

of obese ZDF rats compared to their lean counterparts. ZDF rats were also used for determining protein expression of drug-metabolizing enzymes in pre-diabetic and diabetic animals. Suh et al.²³ reported significantly decreased mRNA expression of the *Cyp1a2* gene in diabetic animals. On the other hand, mRNA expression of the *Cyp1a2* gene in pre-diabetic animals was increased.

Similar to db/db mice, inbred Wistar rats are used as a rat model for type 2 diabetes research. Inbred male Wistar rats were used to study whether lipid accumulation in early steatotic liver affects CYP expression. The results showed significantly decreased CYP1A protein expression in rats with hepatic steatosis as compared to controls²⁴. This finding corresponds to results from the studies of db/db mice by Yoshinari et al.²¹ study and of ZDF rats by Suh et al.²³.

The other member of the CYP1A family is the CYP1A1 enzyme which is also active in drug metabolism; it is expressed in the placenta and also in the fetal liver. An interesting study comparing CYP1A1 activity in full-term placentas of obese and non-obese women counterparts was performed by DuBois et al.²⁵. Furthermore, CYP1A1 activity in fetal liver from a primate (non-human) model of maternal obesity was determined in the same study. The microsomal activity of CYP1A1 was significantly decreased in placentas of obese women. On the other hand, cytosolic CYP1A1 activity was not affected. In fetal liver of non-human primates, the same results were found, that is, a decrease in microsomal CYP1A1 activity²⁵.

The results of various studies indicate that obesity may affect CYP1A enzyme levels in the organism. However, many factors play a role, especially the model of obesity that is used.

Subfamily of CYP2A enzymes

The subfamily of CYP2A enzymes in humans includes the CYP2A6, CYP2A7 and CYP2A13 forms¹³. Of those, CYP2A6 is the most important enzyme involved in drug metabolism. It is predominantly expressed in the liver²⁶ and accounts for ~4% of CYP in the human liver¹³. It is responsible for metabolism of about 3% of marketed drugs¹³. The most significant CYP2A6 substrates are coumarin and nicotine^{27,28}. In addition to these substrates, CYP2A6 metabolizes many drugs, toxic compounds or procarcinogens^{13,28,29} (some of them are listed in Table 1). CYP2A6 is also involved in the metabolism of bilirubin as an endogenous substrate³⁰.

Table 1. Typical substrates, inhibitors and inducers of the human CYP2A6 enzyme.

Enzyme	Substrates	Inhibitors	Inducers
	Cotinine ⁵⁸	8-Methoxypsoralen ²⁸	Carbamazepine ⁶¹
	Coumarin ⁵⁹	Tranylcypromine ⁶⁰	Dexamethasone ¹³
A6	Disulfiram ⁵⁸	Tryptamine ⁶⁰	Phenobarbital ⁶²
P2,	Efavirenz ¹³	Menthofurane ⁵⁸	Rifampicin ¹³
CĄ	Halothan ²⁸	Pilocarpine ¹³	
	Nicotine ²⁷		
	Valproic acid ²⁸		

A human *in vitro* study showed significantly decreased CYP2A6 enzyme activity in human fat-overloaded hepatocytes compared with control samples³¹.

On the other hand, an animal study using MSG-treated mice showed significantly increased murine CYP2A5 enzyme activity, protein and mRNA expression when compared to control mice. Coumarin was used as a substrate for determination of CYP2A5 activity in mice ¹⁰. Similarly, Watson et al. reported increased CYP2A activity (testosterone 7α -hydroxylation) in ob/ob mice in comparison with lean mice²⁰.

Subfamily of the CYP2B enzyme

Although the role of the CYP2B6 enzyme in obesity is not significant, it significantly contributes to drug metabolism³². This enzyme metabolizes ~8% of clinically used drugs¹³. Bupropion, pethidine, propofol and ketamine are drugs metabolized by CYP2B6 (ref.³³). It is also partly involved in the metabolism of nicotine³⁴. This enzyme accounts for ~3-6% of the total CYP content in the liver³³.

Watson et al. reported significantly increased CYP2B activity in male *ob/ob* mice compared to their lean counterparts. In their study, pentoxyresorufin was used as a marker of CYP2B activity²⁰. Furthermore, an *in vivo* study of *db/db* mice found significantly increased CYP2B10 protein as well as *Cyp2b10* mRNA levels as compared with a control group²¹. According to Cheng et al., it is very important to take into account gender specificity. Their study showed decreased mRNA expression of the *Cyp2b10* gene in the liver of female *ob/ob* mice as compared with control mice. Conversely, mRNA expression of *Cyp2b10* was increased in male *ob/ob* mice³².

The use of rat models of obesity often leads to results different from those in mouse obesity models. In a study focused on determination of *Cyp2b2* mRNA expression in an obese Zucker rat model³⁵, decreased mRNA levels of this gene in the liver were observed. On the other hand, Zucker rats exhibited alterations in transcription³⁵.

Subfamily of CYP2C enzymes

The subfamily of CYP2C enzymes consists of the following forms: CYP2C8, CYP2C9 and CYP2C19. The members of the CYP2C subfamily account for ~20% of human hepatic CYP content. The CYP2C8 form is not very important in drug metabolism^{13,11}. The latter forms of this subfamily metabolize 10% and 5% of drugs, respectively¹⁹. All forms in the CYP2C subfamily possess genetic polymorphism³⁶.

CYP2C9 is the major enzyme of the CYP2C subfamily in the human liver. This form metabolizes clinically important medications including antidiabetics (e.g. tolbutamide, glipizide), anticonvulsants (e.g. phenytoin), anticoagulants (e.g. warfarin) and several anti-inflammatory drugs (e.g. ibuprofen) (ref. 13,19,36). The anticoagulant drug warfarin exists in two forms (R- and S-enantiomer). The S-enantiomer of warfarin is predominantly metabolized by CYP2C9 (ref. 37). Some steroids and arachidonic acid are endogenous compounds which may also be metabolized by CYP2C9 (ref. 13).

CYP2C19 was the first CYP2C enzyme in which the genetic polymorphism in the metabolism of mephenytoin was discovered¹³. According to population studies, individuals can be classified as poor or extensive metabolizers of this drug. The S-enantiomer is more rapidly hydroxylated than the R- enantiomer^{13,36}. CYP2C19 metabolizes various therapeutic agents including anticonvulsants (e.g. mephenytoin, felbamate^{36,38}), antidepressants (e.g. amitriptyline, imipramine^{13,36,39}), benzodiazepines (e.g. diazepam¹³) and antiulcer drugs (e.g. omeprazole³⁶). CYP2C19 also metabolizes endogenous substrates, such as progesterone and melatonin¹³.

The effect of obesity on CYP2C enzyme activity and expression, using both animal models and humans, was investigated in several studies. A human study showed only slightly increased CYP2C9 activity in obese individuals¹⁹. In the study, the anticonvulsant phenytoin, the anti-inflammatory ibuprofen and the antidiabetics glimepiride and glipizide were used as CYP2C9 substrates. Higher activity was also observed for the CYP2C19 enzyme in obese than in non-obese individuals. Diazepam was used as a CYP2C19 substrate. Interestingly, in the obese group, no changes in CYP2C19 activity were observed when desmethyldiazepam was used as a substrate¹⁹.

Studies with the db/db mouse model reported no significant differences in protein expression of CYP2Cs. On the other hand, the mRNA level of Cyp2c29 was significantly increased in db/db mice compared to a control group²¹. In a study with mice treated with MSG for induction of obesity, no significant changes in CYP2C enzyme activity, protein expression and gene expression were found as compared to control mice¹⁰.

In an *in vivo* experiment with ZDF rats, researchers found significantly decreased expression of the *Cyp2c39* gene in 12-week-old ZDF rats. *Cyp2c39* gene expression was increased in 6-week-old ZDF rats. This suggests that the liver of 12-week-old ZDF rats has an impaired ability to manage oxidative stress and certain xenobiotics²³. Zhang et al. reported significantly decreased protein expression of rat hepatic microsomal CYP2C11 in early steatosis²⁴.

Subfamily of the CYP2D enzyme

There is a single protein-coding gene in the human subfamily CYP2D named CYP2D6 (ref. 13). This enzyme is expressed in the human liver and represents only about 4% of total CYP (ref. 16). CYP2D6 has been characterized as a typically polymorphic drug-metabolizing enzyme 40. The genetic polymorphism causes the presence of three major phenotypes of oxidative metabolism divided into the following groups: slow, rapid and ultrarapid metabolizers (from null alleles to multiple gene amplification) (ref. 13,11). This enzyme metabolizes about 20% of therapeutic agents 13, such as antidepressants (e.g. nortriptyline, paroxetine, fluoxetine), neuroleptics (e.g. haloperidol, zuclopenthixol), antiarrhythmics (e.g. propafenone, mexiletine), β-adrenoreceptor blockers (metoprolol, propranolol, bufuralol) (ref. 13,16,40) and others.

There are only a few studies dealing with the influence

of obesity on CYP2D6 expression and activity. Donato et al.³¹ studied the *in vitro* effect of lipid accumulation on various CYP enzymes including CYP2D6. Human hepatocytes were incubated with long chain free fatty acids⁴¹ to induce lipid accumulation; a decreased CYP2D6 enzyme activity was observed. The decrease in CYP2D6 activity in human hepatocytes with excess fat may be due to a decreased mRNA level of this gene. On the contrary, Cheymol et al.⁴² showed increased CYP2D6 activity in obese subjects; nebivolol was used as a CYP2D6 substrate.

An *in vivo* study showed no differences in CYP2D enzyme activity, protein and mRNA expression between normal mice and MSG-obese animals¹⁰. The results of individual studies vary; therefore, further studies are needed to confirm or refute the changes found in CYP2D expression and activity.

Subfamily of the CYP2E enzyme

The mammalian subfamily CYP2E only includes the CYP2E1 gene. The human CYP2E1 enzyme form constitutes 3% of the total hepatic content of CYP. This enzyme is known to take part especially in the metabolism of ethanol^{13,11}. Patients addicted to alcohol may be at increased risk of hepatotoxicity of acetaminophen (paracetamol) because of CYP2E1 induction by alcohol, leading to production of a reactive, toxic compound⁴³. In addition to the metabolism of ethanol as a CYP2E1 inducer, this enzyme is also involved in the metabolism of acetone and other known CYP2E1 inducers^{13,11}. Prolonged exposure to compounds (in large amounts) that induce this single form of the CYP2E subfamily may cause formation of free radicals, lipid peroxidation and liver damage⁷. The examples of CYP2E1 inhibitors, inducers and substrates are listed in Table 2. The CYP2E1 enzyme is involved in the metabolism of only about 5% of drugs on the market.

Despite lower mRNA expression of *Cyp2e1* and its limited involvement in drug metabolism, the effect of obesity on CYP2E1 activity has been described in several studies¹⁹. Obesity is considered to be one of the inducers of the CYP2E1 enzyme³⁷.

A common comorbidity of obesity is nonalcoholic fatty liver disease; therefore, the hepatic metabolism via CYP2E1 is very important in obesity⁷. Most studies of both human and animal obesity demonstrated increased activity and expression at protein and mRNA levels of the CYP2E1 enzyme. As a selective probe, chlorzoxazone is

widely used to study the impact of obesity on CYP2E1 activity^{10,44,45}.

A study of obesity using human liver microsomal fractions prepared from diabetic and demographically matched nondiabetic donors observed enhanced CYP2E1 activity in diabetic fractions⁴⁴. Also, an increased CYP2E1 activity (using chlorzoxazone as a probe substrate) was observed in obese patients in comparison with non-obese ones¹⁹.

In an animal study using mice with MSG-induced obesity, significantly increased activity and protein expression of hepatic CYP2E1 in obese mice was found. Also, in the small intestine and colon of obese mice, increased expression of *Cyp2e1* compared to their lean counterparts was observed¹⁰.

Conversely, significantly decreased activity of CYP2E1 was observed in *ob/ob* mice²⁰. Gene expression of *Cyp2e1* was decreased in the liver of female as well as male ob/obmice. Decreased mRNA expression of the Cyp2e1 gene in male ob/ob mice was in accordance with protein expression. However, protein expression of CYP2E1 in female ob/ob mice liver did not differ between obese and control mice³². In another study with ob/ob mice, significantly decreased CYP2E1 expression of protein and mRNA was observed in females; males also exhibited decreased expression but not significantly⁴⁶. Significantly decreased CYP2E1 enzyme activity was seen in the liver of ZDF rats as well as in ob/ob mice of both genders. In male ZDF rats, significantly decreased protein expression and non-significantly decreased mRNA expression were observed⁴⁶. No significant changes in CYP2E1 protein expression as well as in expression of the Cyp2e1 gene in db/db mice were found²¹.

The induction of CYP2E1 in the liver and fat was investigated in three groups of Zucker rats: normal Zucker rats fed with a normal diet (ND), normal Zucker rats fed with a high-fat diet (HF) and genetically obese Zucker rats fed with a normal diet (OB). Higher induction of the CYP2E1 protein in both liver and fat was observed in OB rats compared to HF rats. CYP2E1 activity was also increased in OB rats. In the liver, slightly higher mRNA expression of *Cyp2e1* in HF and OB rats compared to ND rats was observed. In the fat, significantly higher mRNA expression of CYP2E1 in HF and OB rats compared to ND rats was seen⁴⁵. An exception to rat obesity models was a study by Zhang et al. where significantly decreased protein expression of CYP2E1 was observed in male

Table 2. Examples of substrates, inhibitors and inducers of the human CYP2E1 enzyme.

Enzyme	Substrates	Inhibitors	Inducers	
CYP2E1	Acetaminophen ⁴³ Chlorzoxazone ⁵⁹ Benzene ⁶³ Clozapine ²⁴	4-Methylpyrazole ⁶⁴ DEDTC* ⁶² Disulfiram ⁴³ Tetrahydrofurane ⁶² Watercress ¹⁷	Acetone ¹¹ Ethanol ⁶² Isoniazid ⁶² Tobacco ¹⁷	

^{*} DEDTC - N,N-diethyldithiocarbamate

Wistar rats. Clozapine was used as a probe for CYP2E1 activity and the study reported decreased activity of the CYP2E1 enzyme²⁴.

Subfamily of CYP3A enzymes

The CYP3A subfamily in humans consists of the following four CYP genes: CYP3A4, CYP3A5, CYP3A7 and CYP3A43 (ref.13). CYP3A4 and CYP3A5 are the most abundant enzymes located in the human liver and gastrointestinal tract during adult life. On the other hand, in the fetal liver and uterine endometrium, the CYP3A7 enzyme is expressed. However, the amount of this enzyme rapidly decreases in the first weeks of life⁴⁷. Finally, the function of the hepatic CYP3A43 enzyme is not yet known^{44,48}. The CYP3A4 enzyme is the most important CYP form essential for detoxification of xenobiotics and metabolism of most drugs¹¹. This form constitutes about 30% of liver CYP (ref. 13). It is also responsible for several clinically significant drug-drug interactions (e.g. interactions of statins, azoles, antivirals) (ref. 17). Its active site is large, adaptable and it can hold and metabolize many lipophilic compounds, such as immunosuppressants (e.g. cyclosporine A, tacrolimus), anticancer drugs (e.g. taxol) and many others¹³. There are studies reporting the effect of obesity on the CYP3A enzyme. There are a few specific substrates for determination of enzyme activity of CYP3A, such as testosterone¹⁰ or midazolam⁴⁴. Additional specific substrates of the CYP3A4 enzyme are summarized in Table 3.

With regard to the effect of diabetes on CYP activity and expression, there are differences between humans and animals. In fact, these may stem from several factors, such as gender, age, degree of diabetes or duration of the disease⁴⁹. One of the studies dealing with the influence of obesity on CYP enzyme activity and expression reported a significant decrease in CYP3A4 enzyme activity and

protein level in human liver microsomal fractions from diabetic donors. This study indicates that diabetes is associated with a significant reduction in liver CYP3A4 enzyme activity and protein expression⁴⁴.

Brill et al.¹⁹ reported results of a clinical study showing the values of the clearance of CYP3A4 substrates in obese and non-obese patients. The authors found that obesity was associated with significantly lower clearance for most CYP3A4 substrates. Their results correspond with data from an earlier study⁵⁰. Clearance of N-methylerythromycin and triazolam as CYP3A4 probes⁵¹ was significantly lower in obese patients compared to non-obese ones. Also alprazolam, cyclosporine and midazolam showed a trend towards decreased clearance values in obese individuals¹⁹. Taranabant, which is metabolized by CYP3A4, is used for weight loss and reduced fat mass; clearance of this drug was also decreased in obese individuals in comparison to their non-obese counterparts⁵². These results are attributed to either reduction in CYP3A enzyme activity or to increased binding to proteins¹⁹.

A study using male ICR mice fed with two different high-fat diet (HFD) types (36% kcal from fat for 5 weeks and 32% kcal from fat for 40 days) reported altered protein and mRNA levels of the most important biotransformation enzyme CYP3A. The experiment demonstrated a significant decrease of CYP3A protein and Cyp3a11 gene expression in the mice liver with nutritionally induced obesity⁵³. Other authors used adult male CD1 mice fed with HFD (60% kcal from fat) as an experimental group and low-fat diet (10% kcal from fat) as a control group (both groups were fed for 14 weeks). Cyp3a11 gene expression was significantly decreased in the HFD mice compared to the control animals. The observed decrease in mRNA level of *Cyp3a11* gene was confirmed by decreased enzyme activity. Midazolam was used as a CYP3A4 substrate⁵⁴. In a study of ob/ob mice, expression of the Cyp3a11 gene

Table 3. Overview of substrates, inhibitors and inducers of the human CYP3A4 enzyme.

Enzyme	Substrates	Inhibitors	Inducers
	Amitriptyline ⁶³	Itraconazole ⁴³	Carbamazepine ⁶²
	Cyclosporine ⁶⁵	Ketoconazole ⁶²	Dexamethasone ⁶²
	Diazepam ⁴³	Fluconazole ⁴³	Phenobarbital ⁶²
	Domperidone ⁶⁶	Erythromycin ⁴³	Phenytoin ⁴³
	Erythromycin ⁶²	TAO*70	Rifabutin ⁴³
	Clarithromycin ⁶⁷	Amiodarone ¹⁷	Rifampicin ⁶²
	Etoposide ⁶³	Cimetidine ¹⁷	
4	Indinavir ⁶⁸	Gestodene ⁶²	
CYP3A4	Lidocaine ⁴³	Grapefruit juice ⁴³	
Χ	Midazolam ⁶²	Omeprazole ¹⁷	
Ð	Nifedipine (and dihydropyridine drugs) ⁶²		
	Phenacetin ⁶³		
	Sildenafil ⁶⁹		
	Simvastatin ⁴³		
	Testosteron ⁵⁹		
	Taxol ¹³		
	Verapamil ⁶³		
	Tacrolimus ¹³		

^{*} TAO - Triacetyloleandomycin

was decreased in females; however, male liver showed only not significant changes in *Cyp3a11* mRNA levels. Conversely, protein expression of CYP3A11 did not differ between female *ob/ob* mice and control animals; in males, however, significantly decreased CYP3A11 protein expression was observed³². The protein and gene expression of CYP3A did not significantly differ between *db/db* mice and C57BL/6 mice (control animals) (ref.²¹). On the contrary, a study that used mice with MSG-induced obesity showed increased CYP3A activity and protein levels in the liver of obese mice¹⁰.

Suh et al. used ZDF rats at 6 weeks as a pre-diabetes model and at 12 weeks as a diabetes model. They observed significantly reduced hepatic CYP3A enzyme activity in 12-week-old ZDF rats compared to Zucker lean controls²³. As seen from the above comparisons, the findings described in various studies show that CYP regulation is highly dependent on the obesity model, tissue used and other characteristics⁵⁵.

DISCUSSION AND CONCLUSION

Obesity is a complex metabolic disorder that has become a global health problem. To a large extent, obesity in humans is associated with lifestyle (e.g. excessive intake of calories) rather than genetic defects. Obese patients usually suffer from obesity-related diseases and they are thus dependent on the concomitant use of several drugs. It is absolutely necessary that patients are adequately informed about drugs and their possible drug-drug interactions and adverse reactions that may occur. Nowadays, there is insufficient data about how obesity can affect enzyme activity and expression of CYP enzymes in an individual; therefore, animal obesity models are used as surrogate models of human obesity.

Obesity not only affects the properties of CYP enzymes but it is also associated with the development of different diseases. Obesity is linked to an increased risk of cardiovascular diseases. The BMI is an important factor for classification of overweight and obesity. A recent study⁵⁶ aimed to determine its effect on postoperative outcomes after vascular surgery in humans. The study showed lower rates of mortality and cardiac and respiratory morbidity in obese patients after vascular surgery in comparison with normal patients. Despite this fact, wound complications were more frequent in these obese patients. As already mentioned, the increased incidence of obesity results from various types of cancer. Recent research was focused on exploring the effect of high-fat diet-induced obesity on the growth of orthotopically and subcutaneously injected xenografts of human colon cancer to mice. It was found that human colon carcinoma grew faster in obese and insulin-resistant animals due to consumption of high-fat diet⁵⁷.

This review summarizes how obesity and obesity-related diseases can affect the properties of CYP enzymes (the main enzymes of drug metabolism), especially in animal models. In studies mentioned in this review, the authors documented that animal models of obesity exhibited al-

tered expression of drug-metabolizing enzymes. Several animal models have been developed for the obesity research. However, there are differences within an animal species, in gender or induction of obesity. All these factors may influence the regulation of expression of a particular CYP form. This also most likely explains why various models of animal obesity have given different results.

In general, obesity is (in the majority of experiments with human material) associated with a decrease of CYP activities (except for the CYP2C and CYP2E1 enzymes); similar results were obtained for most CYP results in *ob/ob* mice. MSG administered to newborn mice usually leads to an increase in CYP expression.

In other words, it is very important to select the most appropriate animal model, most closely resembling the properties studied in the particular case of human obesity. Although several well-designed animal models have demonstrated the association between obesity and activity and expression of CYP enzymes, a clear explanation of the mechanisms of the influence of obesity on liver drug metabolism in humans is still lacking.

Search strategy and selection criteria

Literature search using the PubMed, Science Direct and Web of Science databases for the years 1984-2016.

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